#### Extension Bulletin • E-2992 • New • May 2007

# MICHIGAN Potato Diseases



# **Fusarium dry rot** Phillip Wharton, Ray Hammerschmidt and William Kirk Department of Plant Pathology, Michigan State University

## Fusarium dry rot

*Fusarium* spp.: *F. sambucinum* Fuckel; *F. solani* (Mart.) Sacc. var. *coeruleum* (Lib. ex Sacc.) (Deuteromycetes, Hyphomycetes)

## Introduction

Fusarium dry rot is one of the most important diseases of potato, affecting tubers in storage and seed pieces after planting. Fusarium dry rot of seed tubers can reduce crop establishment by killing developing potato sprouts, and crop losses can be up to 25 percent; more than 60 percent of tubers can be infected in storage. All the commonly grown potato cultivars in North America are susceptible to the pathogen, although some are less susceptible than others, and several breeding lines have been reported to have a higher degree of resistance to dry rot.

## **Symptoms**

The first symptoms of Fusarium dry rot are usually dark depressions on the surface of the tuber. In large lesions, the skin becomes wrinkled in concentric rings as the underlying dead tissue desiccates (Fig. 1). Internal symptoms are characterized by necrotic



Figure 1. External symptoms of Fusarium dry rot. Dark depressions form on the surface of the tuber, and the skin becomes wrinkled as the underlying tissue desiccates.

areas shaded from light to dark chocolate brown or black. This necrotic tissue is usually dry (hence the name "dry rot") and may develop at an injury such as a cut or bruise. The pathogen enters the tuber, often rotting out the center (Fig. 2). Rotted cavities are often lined with mycelia and spores of various colors from yellow to white to pink (Figs. 2, 3).

Dry rot diagnosis may be complicated by the presence of other tuber pathogens. Soft rot bacteria (*Pectobacterium* spp.) often colonize dry rot lesions, especially when tubers have been stored under conditions of high relative humidity or tuber surfaces are wet. Soft rot bacteria cause a wet, slimy rot, which can rapidly engross the entire tuber and mask the initial dry rot symptoms (Fig. 4). Dry rot may also accompany late blight infection of tubers and be followed by soft rot bacteria, leading to tubers with symptoms of all three diseases (Fig. 5).



Figure 2. Internal dry rot symptoms. Clumps of white to yellow mycelia line a dry necrotic cavity hollowed out from rotted tissue.



Figure 3. Clumps of mycelium and white to pink to yellow sporulating masses form on the surface of dead skin.

Pythium leak and pink rot also cause brown to black internal discoloration of tubers. However, these are wet rots, and tubers exude a clear fluid when squeezed.

#### **Disease cycle**

Fusarium dry rot is caused by several fungal species in the genus *Fusarium*. *Fusarium sambucinum* (teleomorph *Giberella pulicaris*) is the most common pathogen causing dry rot of stored tubers in North America, but other *Fusarium* species are also known to cause dry rot, particularly *F. solani* var. *coeruleum* and *F. avenaceum*. In Michigan, *F. sambucinum* is probably the main causal agent of dry rot, but *F. solani* var. *coeruleum* may also be present.

Fusarium species are common in most soils where potatoes are grown and can survive as resistant spores free in the soil for very long periods of time. There are two main opportunities in the potato crop cycle for Fusarium species to infect potato tubers in the spring and in the fall (Fig. 7). Fusarium sambucinum and F. solani are commonly found on seed tubers in the spring. Potato seed tubers are maintained in storage at 37°F, which is approximately the temperature at which F. sambucinum is dormant, and consequently, there is minimal development of dry rot in storage. However, some level of Fusarium dry rot is almost always present in commercially available seed. During the preplanting phase of potato production, seed tubers are warmed to about 54°F, then cut into seed pieces prior to planting. Tubers infected with *F. sambucinum* are particularly susceptible to the development of seed piece decay during this phase. In cases of severe disease, seed pieces may rot completely before planting. Alterna-



Figure 4. Dry rot is often followed by bacterial soft rot, which makes the tissue soft, mushy and slimy.

tively, after planting, more than 50 percent of sprouts developing on infected tubers may become diseased and killed outright before emergence. Damage at this stage results in delayed or non-emergence and is usually expressed as poor and uneven stands with weakened plants (Fig. 6). Reduction in crop vigor then results from expenditure of seed energy used to produce secondary or tertiary sprouts to compensate for damage to primary sprouts.

Progeny tubers may become contaminated with *Fusarium* spores as they develop in the late summer and early fall. Tubers are not usually infected until harvest because the pathogen cannot cause infection unless the potato skin is ruptured, which rarely occurs during the growing season. Wounds caused during harvest and handling provide dormant spores on the tuber surface with multiple points of entry into the tuber. Once the pathogen has penetrated the tuber skin, it begins to grow in the tuber tissue, causing dry rot lesions at the point of entry (Fig. 2). In storage, dry rot develops most rapidly at high relative humidity and temperatures of 60° to 70°F. Lower humidity and temperatures retard infection and disease development. However, dry rot may continue to develop at the lowest temperatures safe for storage of potatoes. Young tubers appear to have some resistance to dry rot, which slows disease. Dry rot progresses noticeably faster during the latter half of the storage season.

#### Monitoring and control

From about 1970 to 1985, control of dry rot was primarily and effectively achieved by the postharvest application of thiabendazole (Mertect). However, from the late 1980s onwards, many strains of *F. sambucinum* 



Figure 5. Dry rot (F) may also accompany late blight infection (LB), followed by soft rot bacteria (SR).

became resistant to the benzimidazole fungicides, such as thiabendazole and thiophanate-methyl, resulting in poor control of dry rot. The pathogen infects through wounds; therefore, modification of tuber handling to reduce wounding during harvest and storage and the use of an effective seed treatment in combination with good management practices during cutting and storage of cut seed before planting are essential to reducing Fusarium dry rot.

## **Cultural control**

Some level of Fusarium dry rot is almost always present in commercially available seed. Even though it is not possible at present to be 100 percent sure that a seed lot is completely free of dry rot, it is sensible to plant seed that meets established seed certification standards. Practicing the following procedures will help prevent dry rot:

- Plant only certified seed. It is critical to purchase seed with as little dry rot as possible, so always inspect seed carefully upon receipt.
- After careful unloading, seed should be stored at 40° to 42°F and 85 to 90 percent relative humidity, and kept ventilated.
- Warm seed tubers to at least 50°F before handling and cutting to minimize injury and promote rapid healing.
- Clean and disinfect seed storage facilities thoroughly before receiving seed.
- Disinfect seed cutting and handling equipment often, and make sure cutters are sharp to ensure a smooth cut that heals easily.
- Do not store seed near a potential source of inoculum (e.g., cull piles).



Figure 6. Seed piece decay and sprout rot lead to poor and uneven stands with weakened plants.

- Treat cut seed with a seed treatment to control seed piece decay and sprout rot (see current recommendations for specific fungicides below).
- Plant seed that has a *Fusarium* problem in warm, well-drained soil to encourage rapid sprout growth and emergence, and lessen the chance for infection.
- In the fall, harvest tubers after their skins have set and when their core temperature is greater than 50°F.
- Monitor stored tubers often for dry rot. Grade out rotten tubers when tubers are removed from storage for marketing.

## **Biological control**

Currently, no commercially available biological control products are registered for the control of Fusarium dry rot. However, studies are under way at MSU to evaluate the use of biofungicides containing the biological control bacteria *Bacillus subtilis* (Serenade, AgraQuest) and *B. pumilus* (Sonata, AgraQuest), and the biocontrol fungus *Trichoderma harzianum* (T-22 Planter Box, Bioworks) for control of Fusarium dry rot on potato. These compounds are being evaluated both as seed treatments and for postharvest application in storage.

## **Chemical control**

### Seed treatment

Several products have been developed specifically for control of seed-borne potato diseases (Table 1) and offer broad-spectrum control for Fusarium dry rot, *Rhizoctonia*, silver scurf and, to some extent, black dot (*Colletotrichum coccodes*). These include Tops MZ, Maxim MZ (and other Maxim formulations + mancozeb) and Moncoat MZ. The general impact of





these seed treatments is marked by improved plant stand and crop vigor, but occasionally, application of seed treatments in combination with cold and wet soils can result in delayed emergence. The delay is generally transient, and the crop normally compensates. The additional benefit of the inclusion of mancozeb is for prevention of seed-borne late blight.

Studies at MSU have shown that the most effective control of Fusarium dry rot is achieved by the application of an effective fungicide, such as fludioxinil (Maxim-based products), prior to planting. Treatment of infected seed pieces with Maxim MZ at 10, 5 or 2 days before planting significantly reduced the percentage of diseased sprouts per tuber and significantly reduced seed piece decay in the varieties Pike and FL1879. Although it may not seem cost-effective to apply seed treatments to healthy seed, these results suggest that applying a seed treatment up to 10 days prior to planting can provide effective control of dry rot and increase rate of emergence, rate of canopy closure and final plant stand. **Postharvest fungicides** 

Although largely ineffective, thiabendazole remains registered for postharvest use on tubers. Few alternative compounds are available for potato tuber treatment in storage; these include chlorine-based disinfectants such as sodium hypochlorite, calcium hypochlorite and chlorine dioxide. Limited information is available on the effectiveness of chlorine dioxide on potato storage pathogens, and results of some studies have suggested that chlorine dioxide does not provide effective tuber protection against Fusarium dry rot.

Studies are under way at MSU to evaluate several of the new reduced-risk fungicides for use in postharvest applications. These fungicides – Quadris, Phostrol and Oxidate – are all currently registered for foliar application to potatoes in the field. **Table 1.** Product name, active ingredient and FRAC<sup>a</sup> resistance management grouping, type and rate of application, and activity of products currently registered for control of Fusarium dry rot of potatoes.

Product⁵	Active ingredient [Chemical group]	Type of application	Rate of application <sup>°</sup>	Activity against dry rot <sup>d</sup>
MonCoat MZ	flutolanil (1.5%) [7] mancozeb (6%) [M3]	Seed treatment (dry)	0.75 - 1.0 lb/100 lb	+++
Maxim 4FS	fludioxinil (40.3%) [12]	Seed treatment (liq.) (for production of certified seed tubers only)	0.08 - 0.16 fl oz/100 lb	+++
Maxim Potato Seed Protectant	fludioxinil (0.5%) [12]	Seed treatment (dry)	0.5 lb/100 lb	+++
Maxim MZ	fludioxinil (0.5%) [12] mancozeb (6%) [M3]	Seed treatment (dry)	0.5 lb/100 lb	+++
Nubark Maxim	fludioxinil (0.5%) [12]	Seed treatment (dry)	0.5 lb/100 lb	+++
Tops MZ	thiophanate-methyl (2.5%) [1] mancozeb (6%) [M3]	Seed treatment (dry)	0.75 lb/100 lb	+++

<sup>a</sup> Fungicide Resistance Action Committee. See (http://www.frac.info) for more information.

<sup>b</sup> Specific instructions are included on the labels of all of the products, and these must be adhered to.

<sup>c</sup> Rate of application is per acre at 34-inch spacing between rows (in-furrow) and per 100 lb of tuber seed pieces (seed treatment, wet or dry).

<sup>d</sup> + signifies excellent activity against dry rot.

Photos, text editing, design and page layout by P.S. Wharton; illustrations by Marlene Cameron. For more information, please visit: http://www.potatodiseases.org

This publication is part of a new series of bulletins on potato diseases in Michigan. Funding for this publication was provided by Project GREEEN, MSU Extension, the Michigan Agricultural Experiment Station and the Michigan Potato Industry Commission.



MICHIGAN AGRICULTURAL EXPERIMENT STATION





MSU is an affirmative action equal opportunity employer. Michigan State University Extension programs and materials are open to all without regard to race, color, national origin, gender, religion, age, disability, political beliefs, sexual orientation, marital status, or family status. Issued in furtherance of Extension work in agriculture and home economics, acts of May 8 and June 20, 1914, in cooperation with the U.S. Department of Agriculture. Thomas G. Coon, Extension director, Michigan State University, E. Lansing, MI 48824. Text information and images in this publication are the property of MSU Extension and the authors and are protected by copyright. References to commercial products or trade names do not imply endorsement by MSU Extension or bias against those not mentioned. Copying of any material in this bulletin is prohibited, except for non-commercial educational purposes, such as teaching, scholarship, research, criticism, and commentary. Unless otherwise noted, users who wish to copy and or print text and images from this bulletin for such uses may do so without MSU's express permission, provided that they comply with the following conditions. They must cite the author and source of the content, and none of the content may be altered or modified. Unauthorized commercial publication or exploitation of text or images in this bulletin is specifically prohibited. Anyone wishing to use this bulletin or any of the images for commercial use, publication, or any purpose other than fair use as defined by law, must request and receive prior written permission from the authors or Michigan State University. © 2007 Michigan State University, All Rights Reserved.

#### New 05:07-1.5M-KMF/DP